

# Packing of monodisperse DNA-RecA protein complexes

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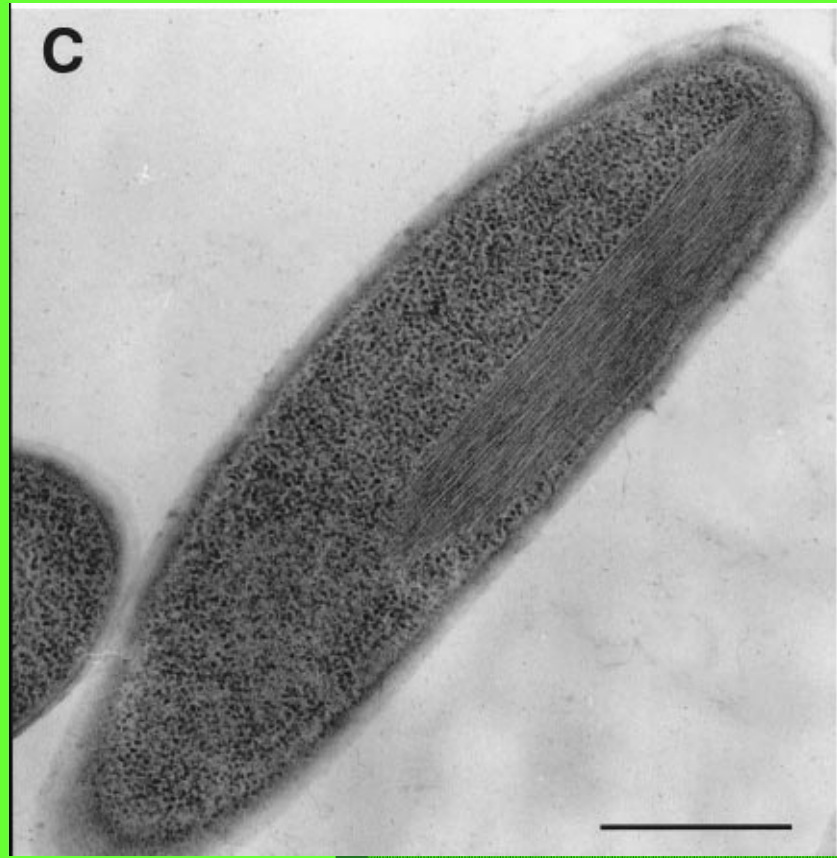
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## Motivation

– Condensed phases are functional structures

- DNA in all living systems: highly condensed and tightly packed (mitotic chromosomes, sperm heads, virus capsids)
- DNA replication, transcription, protection, and repair in highly packed genetic material
- DNA packs by oppositely charged multivalent ions and proteins  
*Mangenot et al. BPJ2003*  
*Raspud et al. PRL 2000*  
*Leforestier & Livolant, BPJ1993*

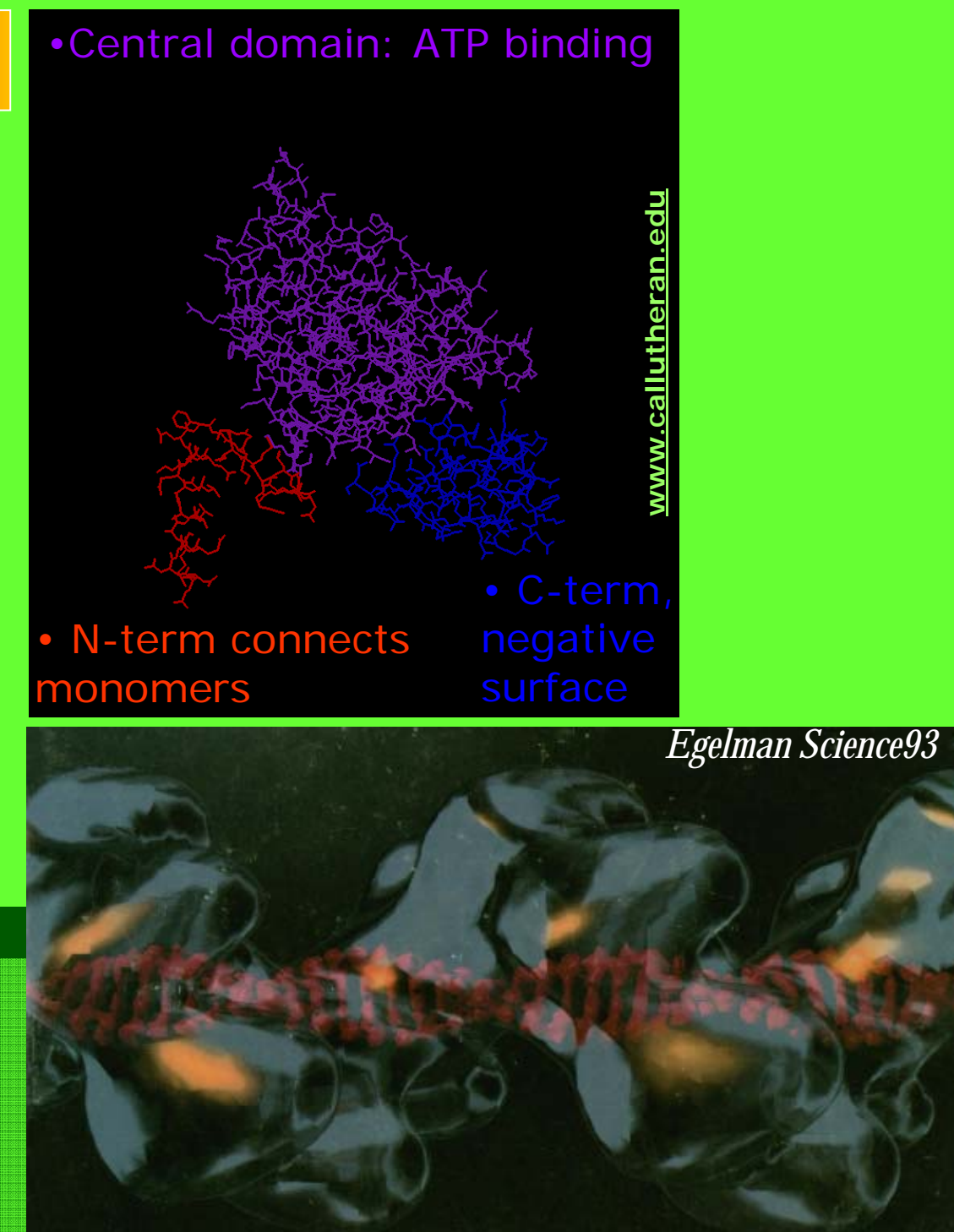


A different packing: intracellular coaggregation of RecA protein and DNA into a macroscopic assembly

*Levin-Zaidman et al., PNAS 2000*

## RecA protein overview

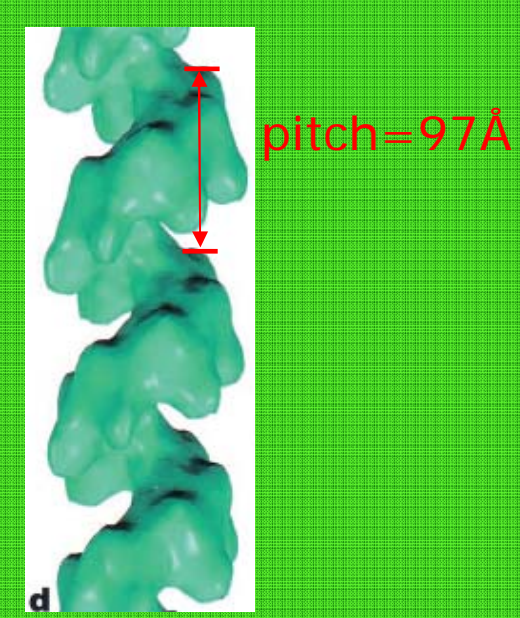
- E.coli RecA
- MW = 37,842
- 352 amino acid residues
- polymerizes in helices around DNA
- Structure → Function
- Archaea and Eukaryota have homologous proteins
- multirole:
  - SOS response LexA repressor cleavage
  - Promotes homologous genetic recombination via DNA strand exchange by forming nucleoprotein filament with free ssDNA ends



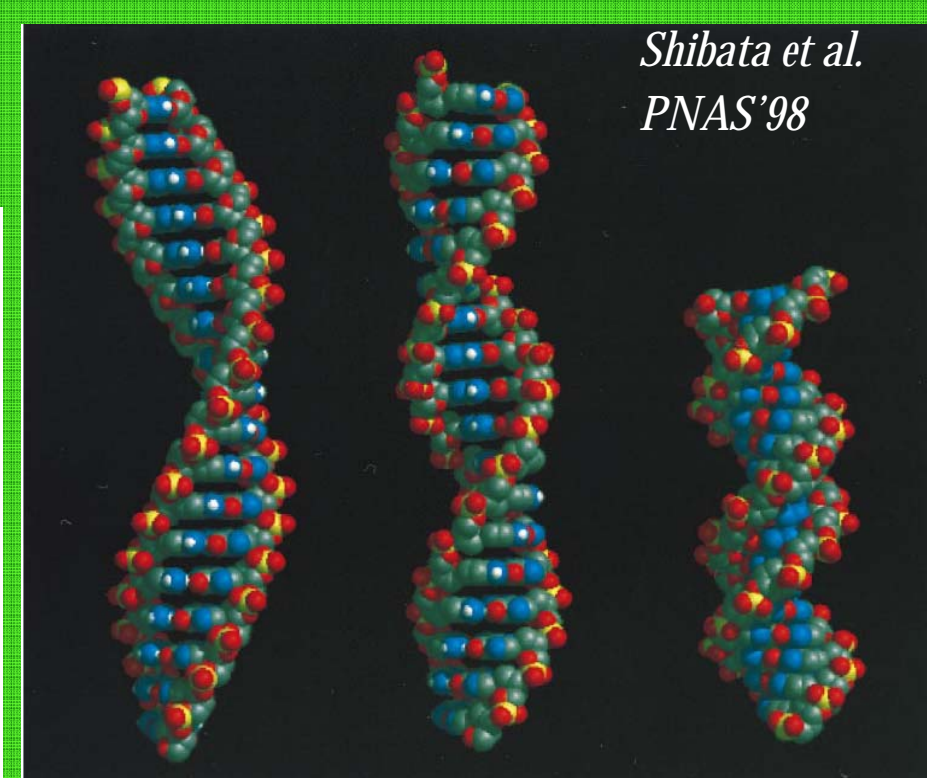
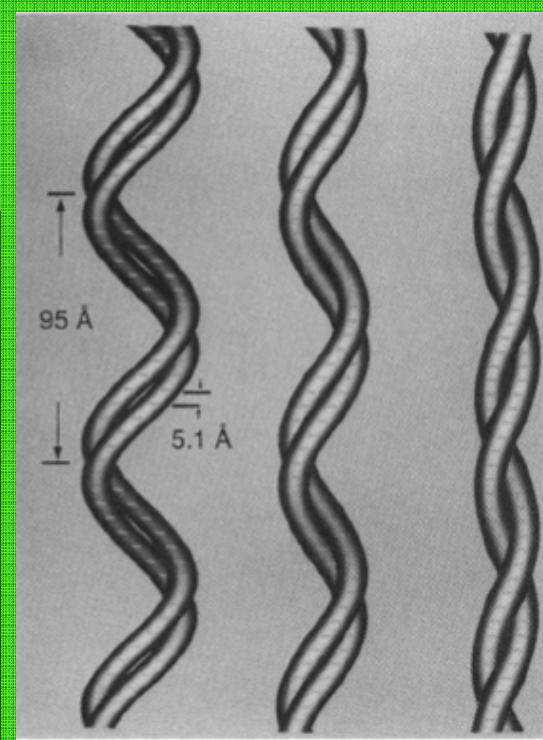
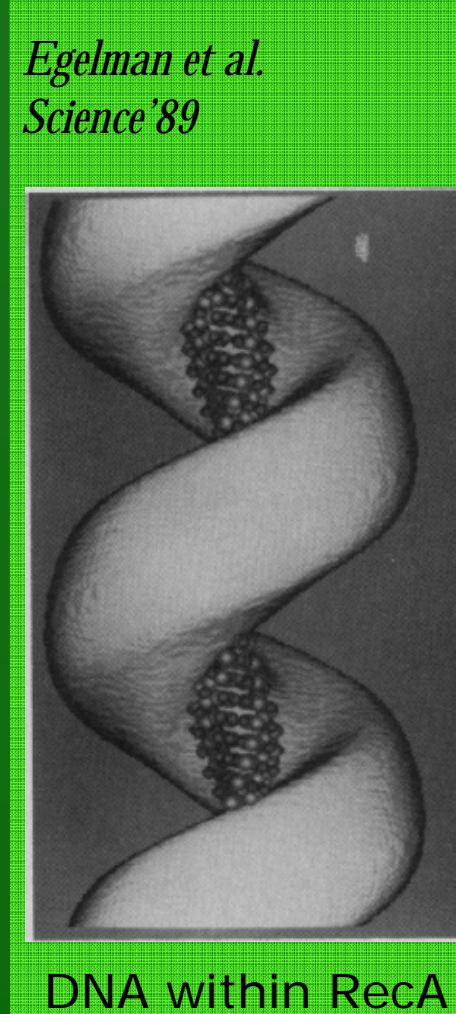
## Samples & materials

- E. Coli RecA protein: overexpression plasmid pAIR79 in E. Coli strain STL327, described in *Lovett, PNAS1989*
  - RecA purification according to *Cox et al. JBC81*
  - 146bp DNA fragments of calf-thymus chromatin, enzymatically digested according to *Strzelecka & Rill JACS1987*
  - ATP-γ-S, Roche
- All concentrations determined by spectrophotometry

## dsDNA within RecA filament



RecA + dsDNA +ATPγS+Mg<sup>++</sup>  
in vivo & in vitro active nucleoprotein filament  
pitch=97Å  
monomers/turn=6.20  
stoichiometry=3bp/monomer



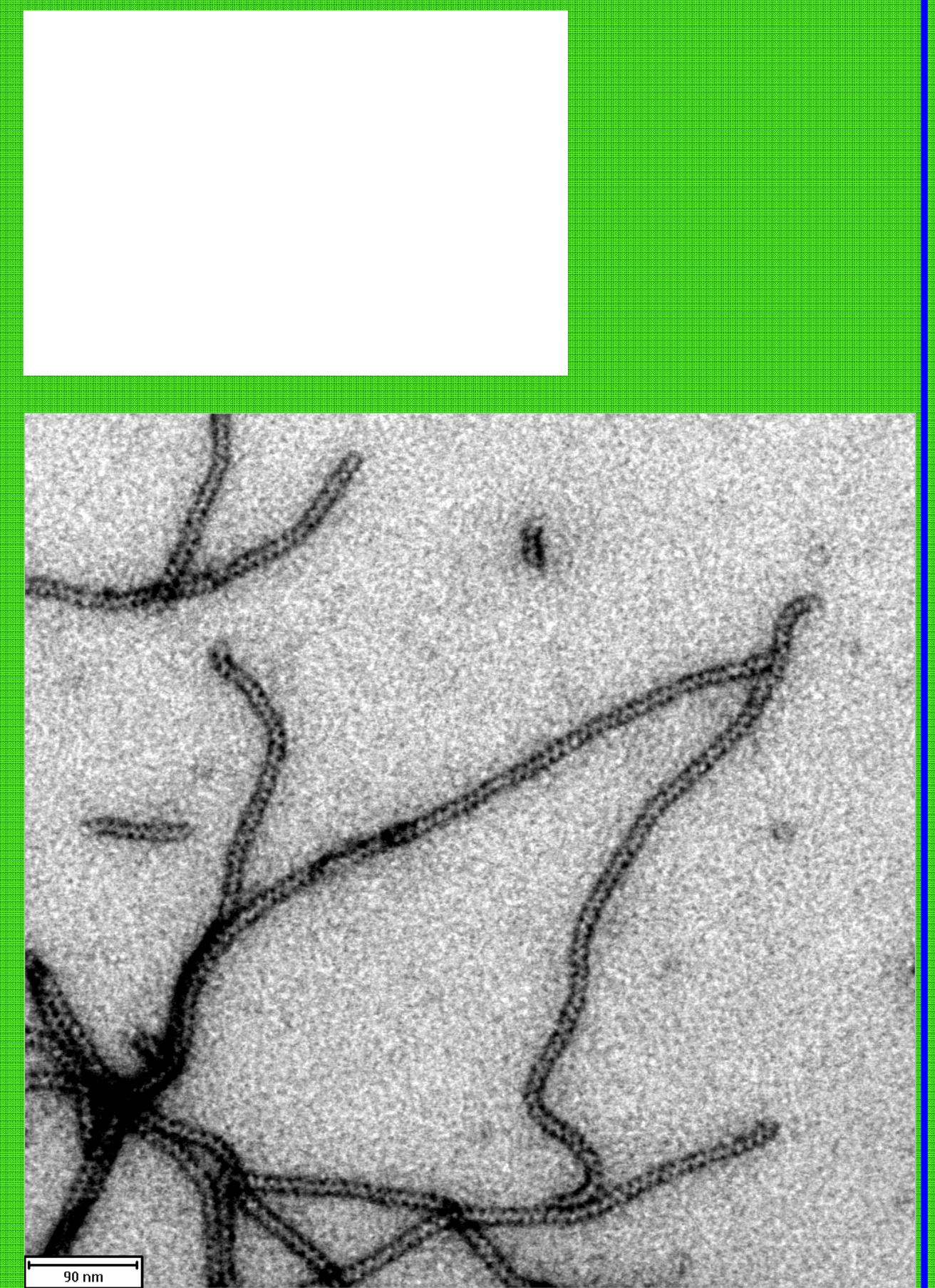
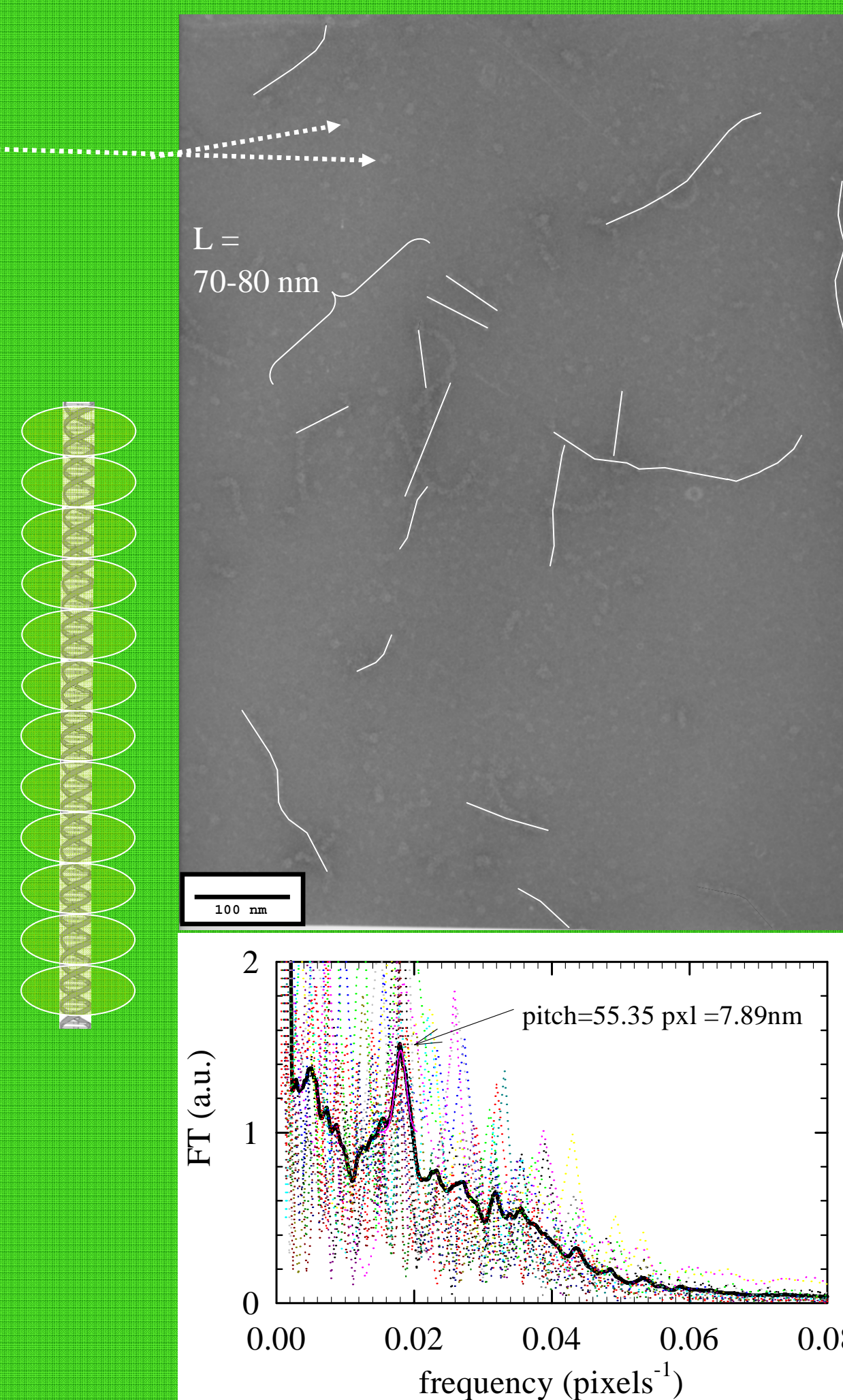
DNA base pair rise: 5.1 Å, 5.1 Å, 3.4 Å  
DNA pitch: 99 Å, 67 Å, 34 Å  
stoichiometry: 3bp/mer, 2bp/mer ?!

• How the models correlate with known parameters of RecA+DNA complex, pitch, stoichiometry, monomers per turn?

## RecA polymers

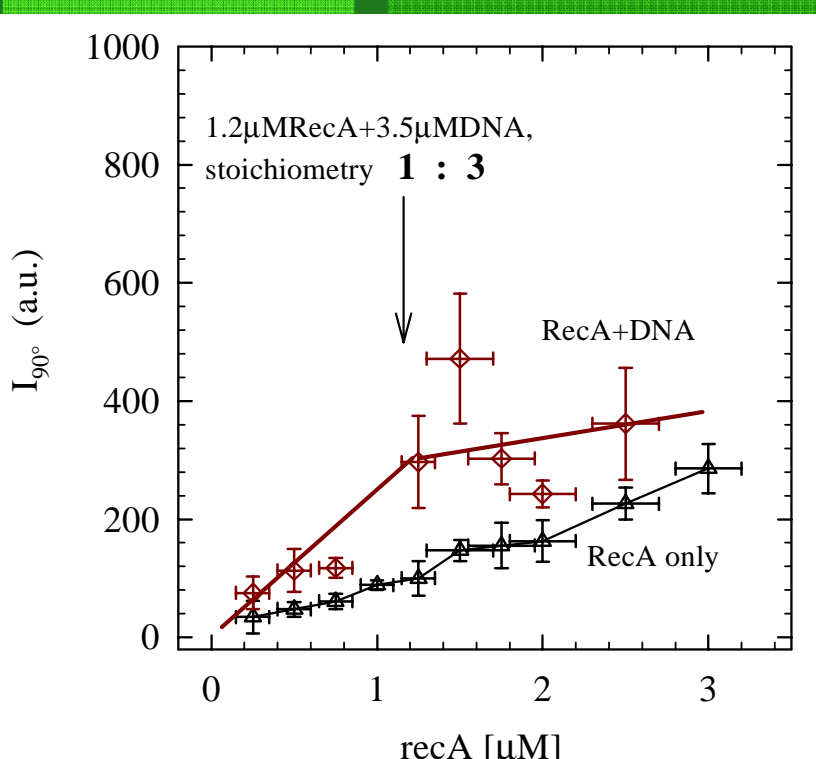
- RecA selfpolymers have similar structure as the complexes  
*Hegner et al. PNAS'99*
- selfpolymers
- bundles, aggregates, crystals
- RecA selfpolymers and aggregates are not intermediates en route to nucleoprotein filaments  
*Morrice & Cox Biochemistry 1985*

## RecA complexation with 146 bp DNA fragments: rodlike nucleoprotein particles and some RecA polymers?

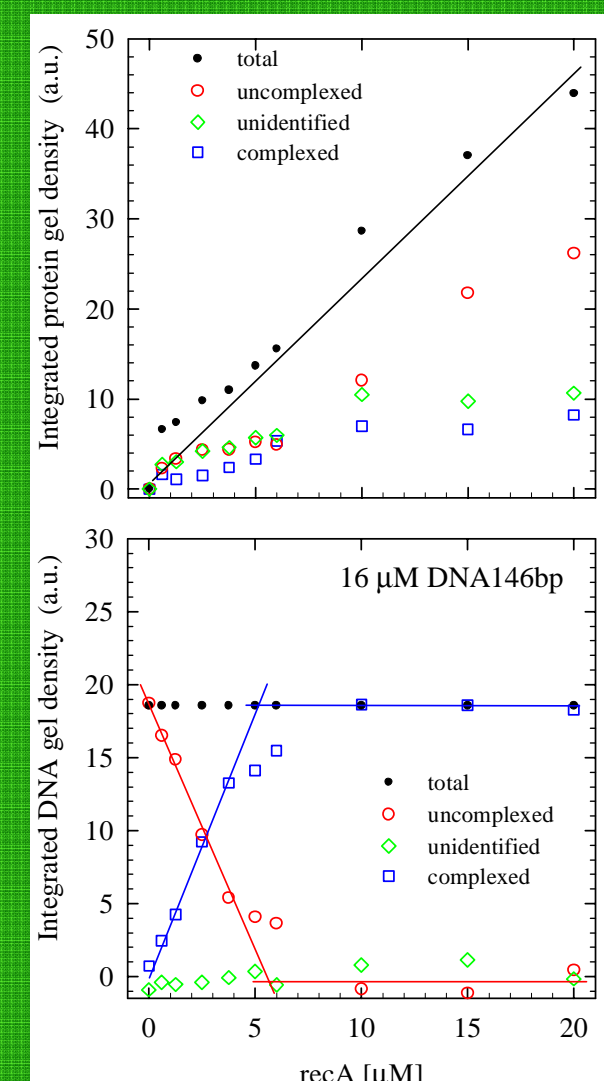


## RecA/DNA stoichiometry: 3/1 or 2/1?

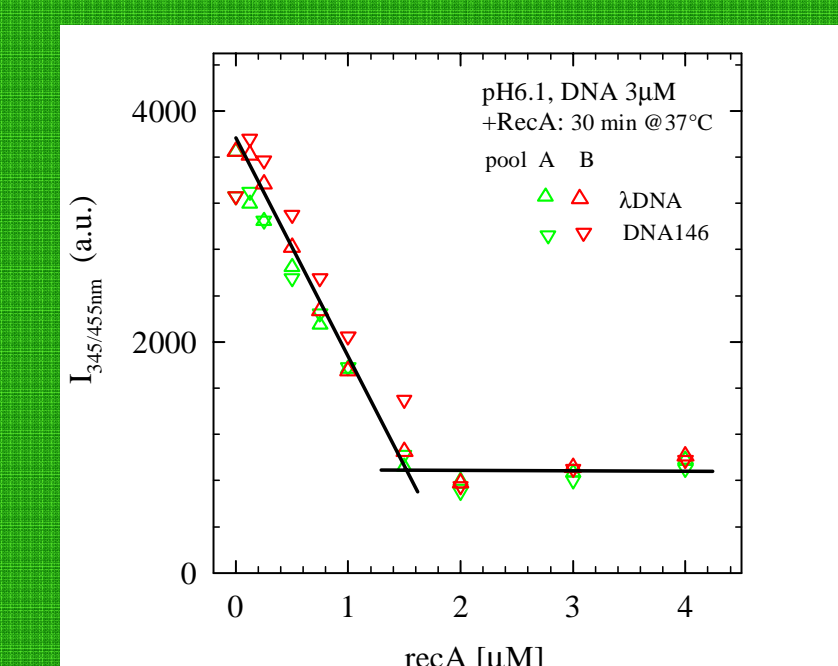
• 90° static light scattering, is it distinguishing RecA polymer from nucleoprotein filaments?



• gel densitometry



• fluorometry – binding of DAPI fluorophore to uncomplexed DNA



## RecA+DNA dense phase formation problem

- besides various liquid crystalline dense phases of DNA only, nucleoprotein dense phase is known, formed by nucleosome core particles and DNA *Mangenot et al. BPJ2003*
- RecA-only filaments & RecA-DNA complexes: bundles and aggregates compete against liquid crystal  
*Egelman & Stasiak, JMB 1988*  
*Di Capua et al. JMB 1982*
- a basic function of the system: DNA strand exchange is initiated by elevated Mg, spermidine or by crowding agents  
*Lavery & Kowalczykowsky, JBC 1992*
- all these also induce aggregation/bundling of RecA based filaments
- DNA liquid crystals form more easily with monodisperse fragmented DNA
- RecA filaments/complexes have similar helical symmetry as DNA *Leforestier & Livolant BPJ 1993*
- Short RecA complexes (formed on short DNA) might organize and not just aggregate

## Summary

The structure of RecA-DNA complex is not solved, and the exact path of DNA within the nucleoprotein filament is not known, although it has been extensively studied by SANS, electron microscopy or NMR  
*DiCapua et al. JMB1990; Yu X. et al. PNAS2001; Nishinaka et al. PNAS1998.*

We form RecA nucleoprotein filaments using very short, monodisperse, 146 bp long DNA. These 75 nm long filaments are shorter than their respective persistence length – i.e. they should behave as straight rods. A system of monodisperse rodlike particles of helicoidal symmetry is capable of forming cholesteric liquid crystalline phase.  
*Leforestier & Livolant, BPJ1993*

Such a dense phase of RecA-DNA filaments might be the most ordered possible preparation, facilitating structural studies.